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and (starting summer 2015)

Core facility bioimaging of the Biomedical Center



Multi-photon Microscopy



Flavors of Multi-Photon-Microscopy

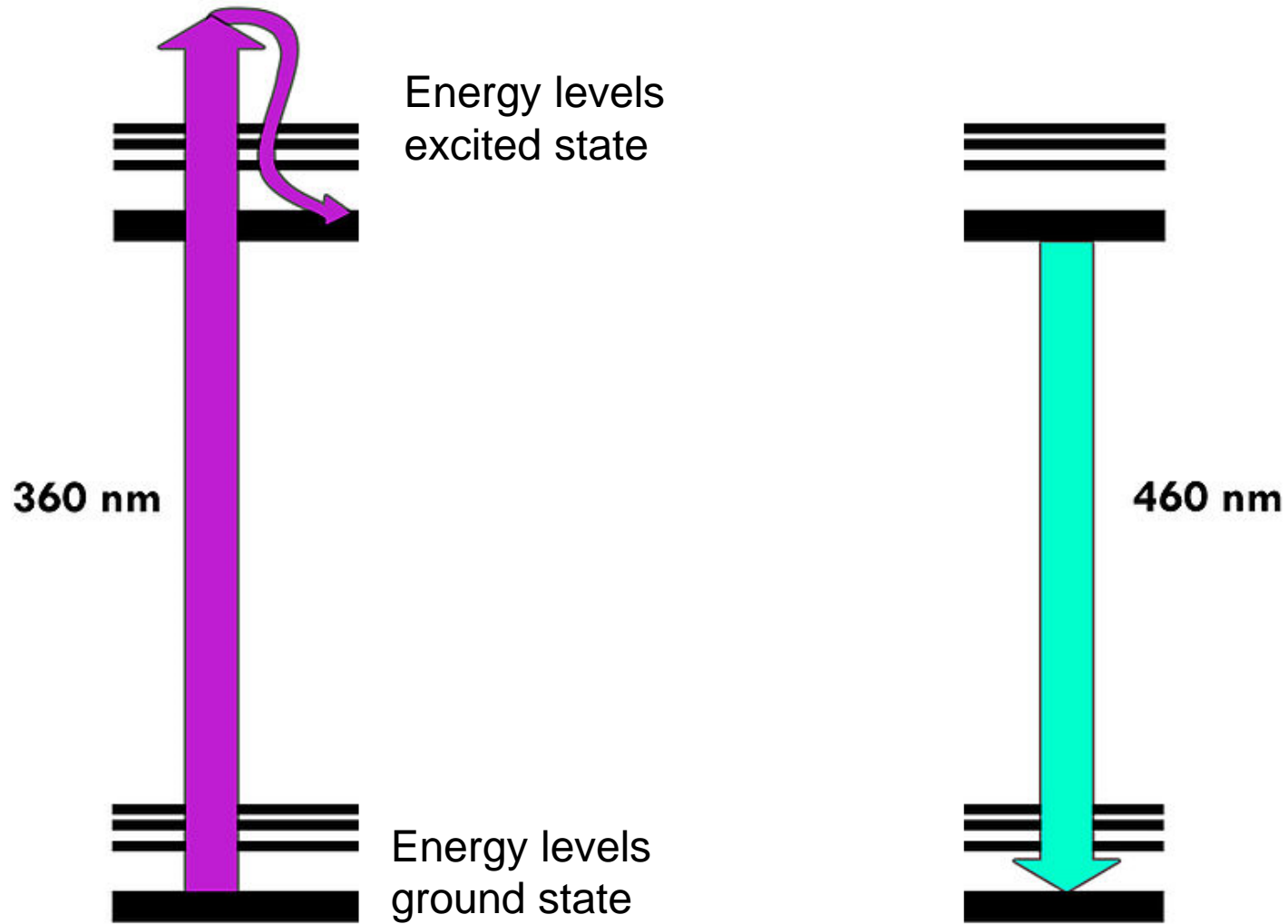
- Two-photon excitation fluorescence (TPEF)
 - Second harmonic generation (SHG)
 - Third harmonic generation (THG)
 - Resonance enhanced THG
 - Plasmon induced luminescence
- Coherent Anti-Stokes Raman (CARS)-Microscopy

Two-photon excitation fluorescence

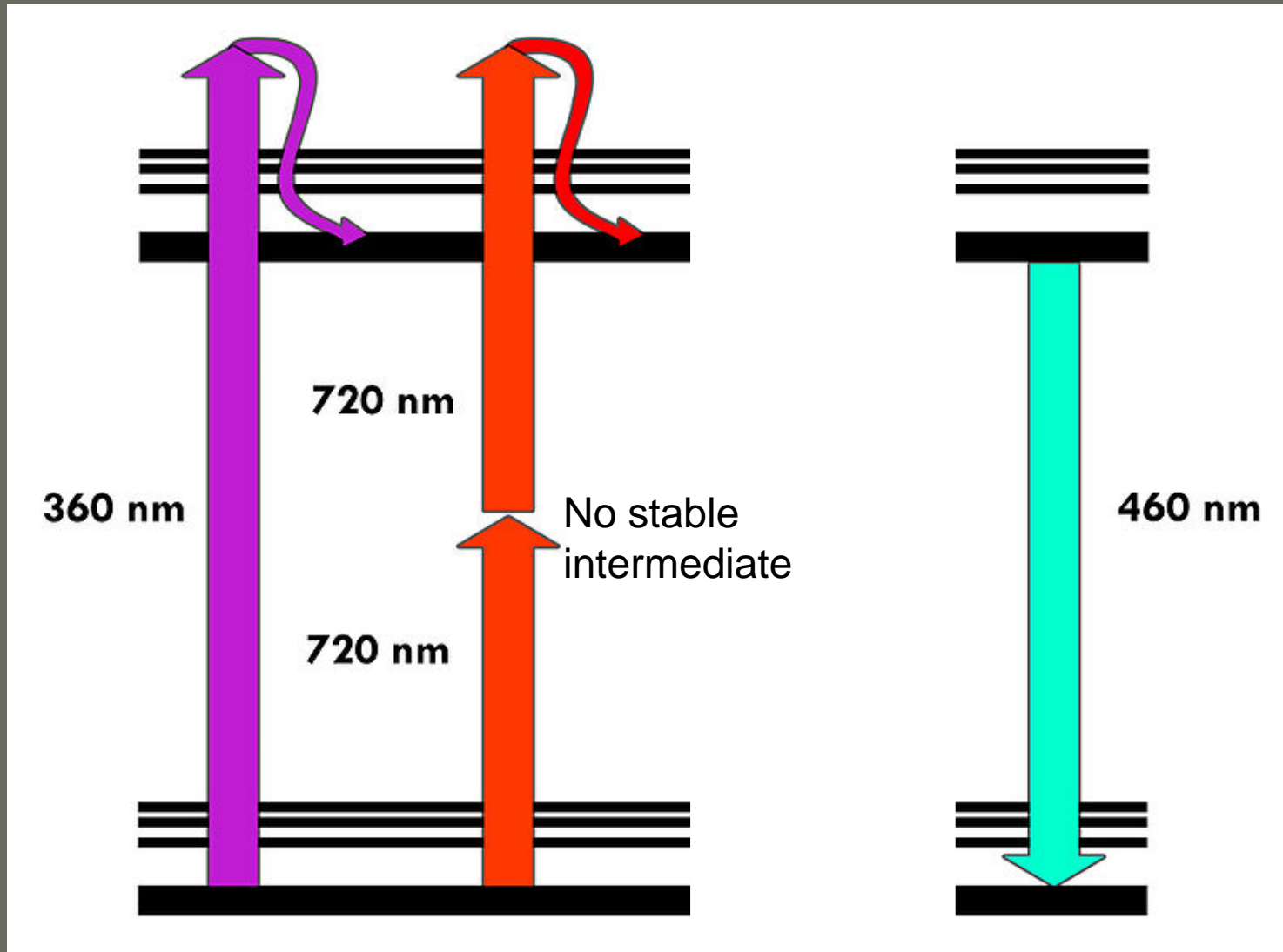
- How
- Why
- What for



1 photon excitation fluorescence



1+2 photon excitation fluorescence

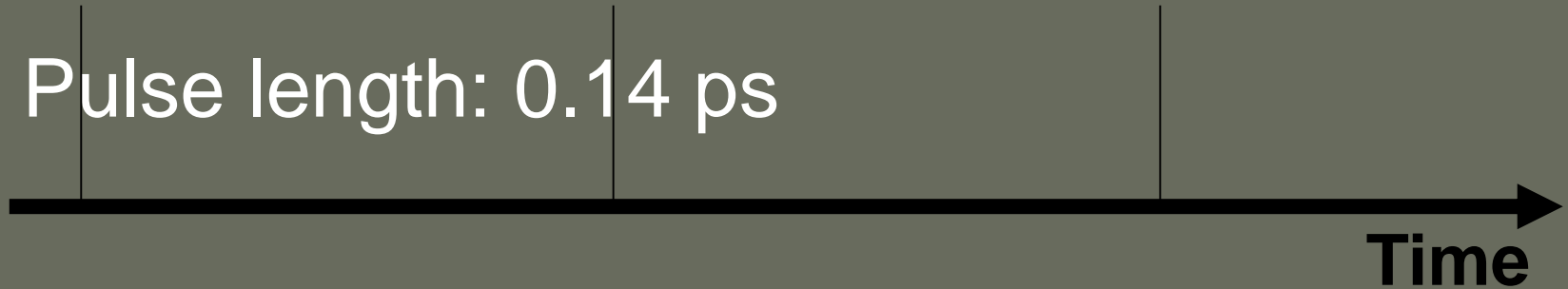


Two-photon excitation...

- ... only works if two photons arrive at the absorbing electron within 1 attosecond, $1 \cdot 10^{-18}$ s (nano, pico, femto, atto).
- This is statistically very unlikely to happen under normal conditions.
- To generate a high enough photon density, a focused, pulsed laser is required.

Pulsed laser (Chameleon Ultra II)

- Pulse length: 0.14 ps



- Pulse interval: 12 500 ps
(= 12.5 ns = 80 MHz)

- So, most of the time, the laser is off, only 0.00112 % on.
- About same relation as a 1 second pulse in one day.

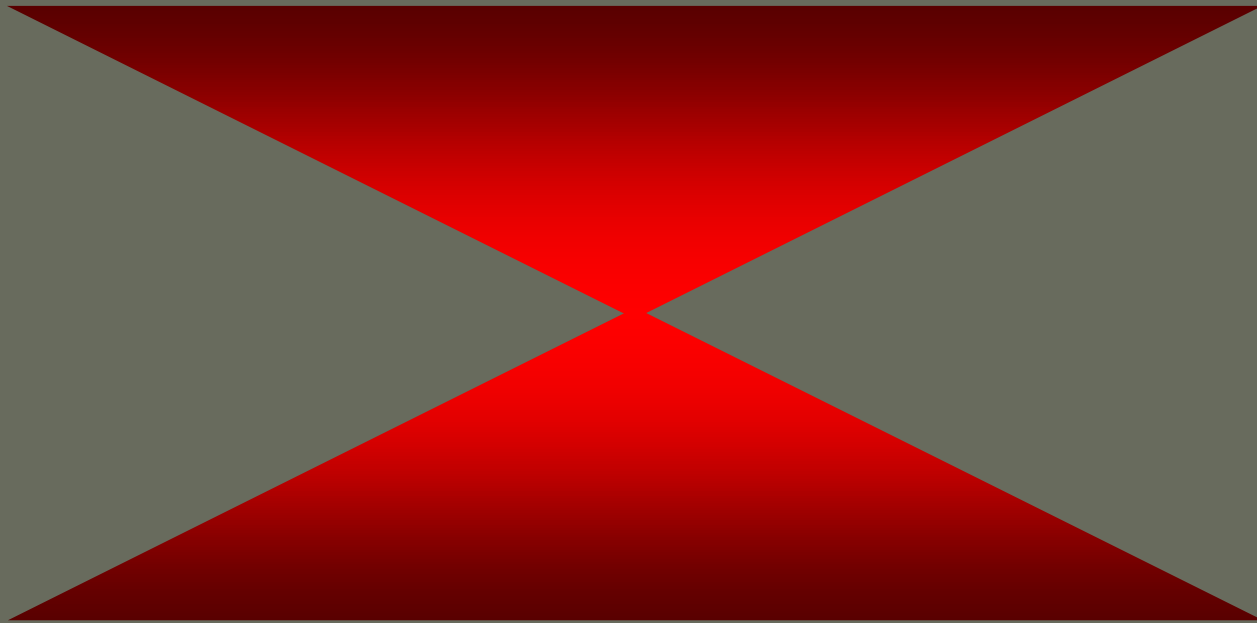
Image formation

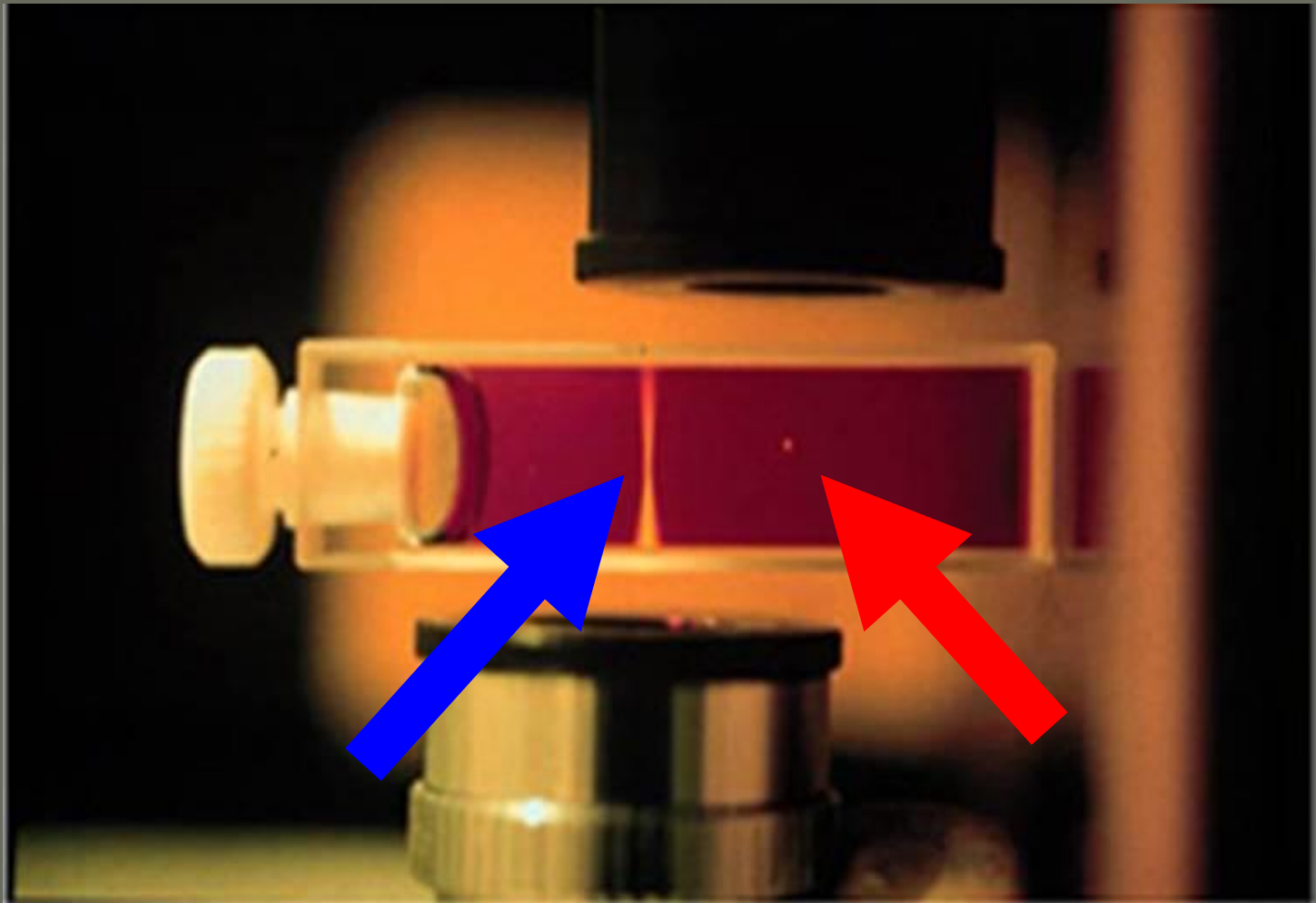
(similar to confocal microscope)

- At any given time, fluorescence comes from only one point.
- This point is scanned over the specimen.
- Detectors („PMTs“) record emitted photons from each point, one by one, the computer constructs the image.

It's complicated and expensive,
so why bother?

- Advantage 1: Excitation and bleaching only at the focal point. Not above and not below.





Advantage 1: No out-of-focus bleaching

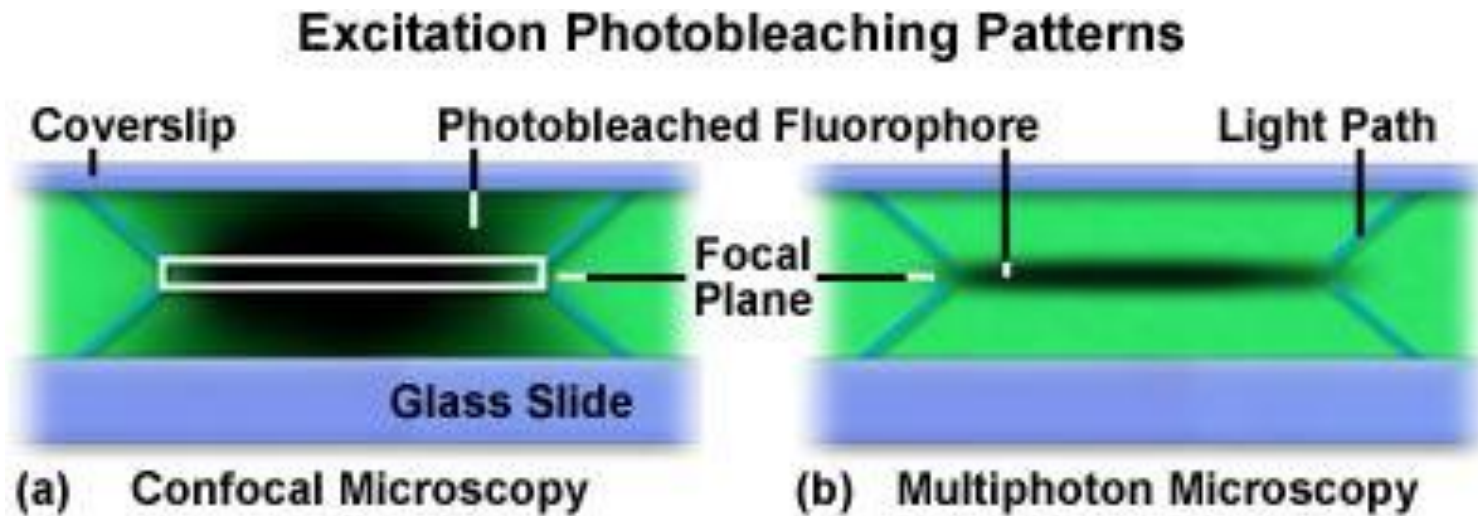


Figure 5

Why bother?

Advantage 2: Deeper penetration, because

- a) scattering decreases with longer wavelengths:
 $1/(\text{nm})^4 \Rightarrow$ factor 16 with doubling of λ

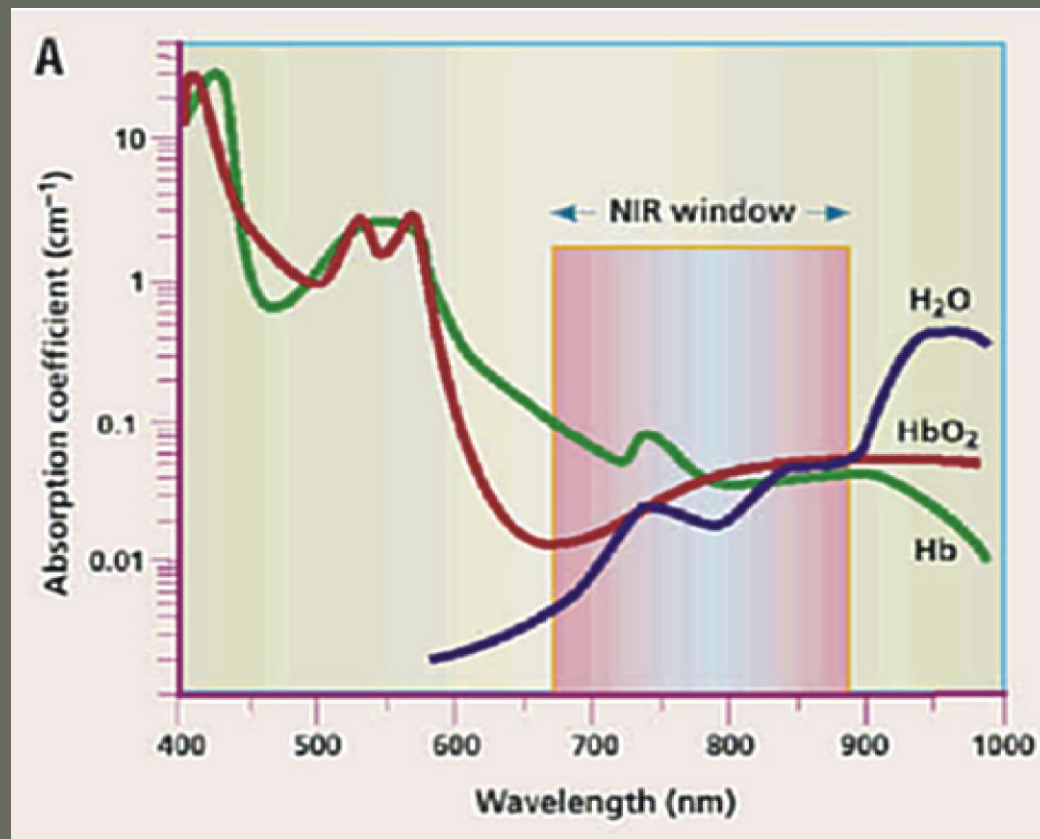


<http://www.foto-fotograf.de/sonnenuntergang1.jpg>

Why bother?

Advantage 2: Deeper penetration, because

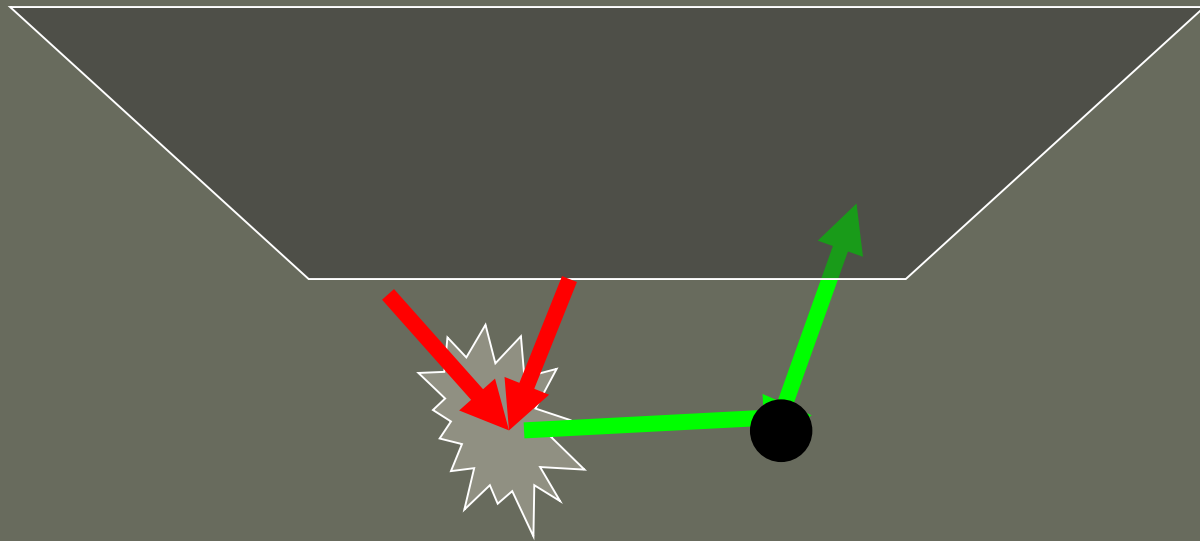
- b) Less absorption of tissue in NIR-range



Why bother?

Advantage 3:

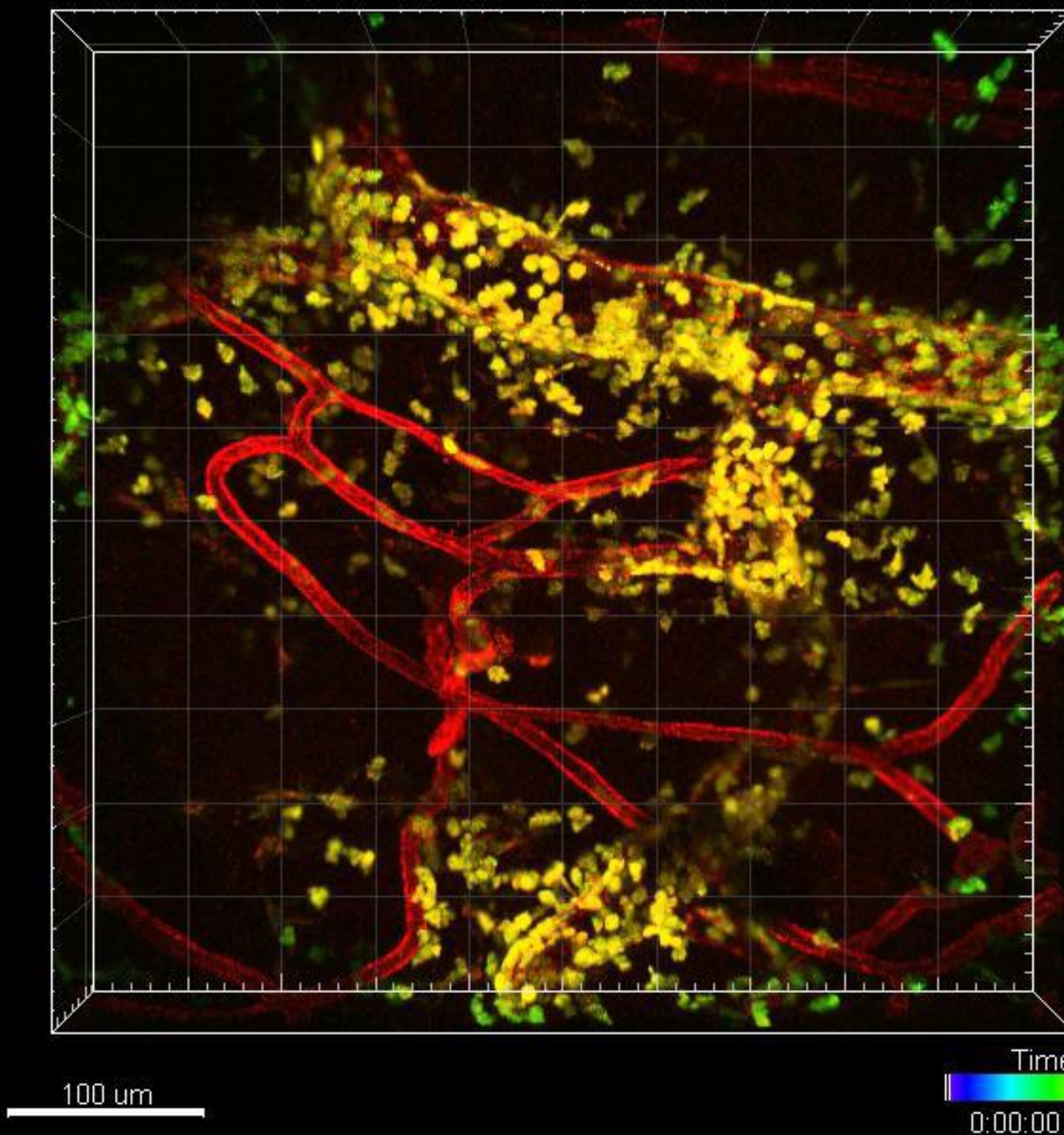
- No loss of emitted photons due to pinhole, non-balistic photons contribute to image



Disadvantages

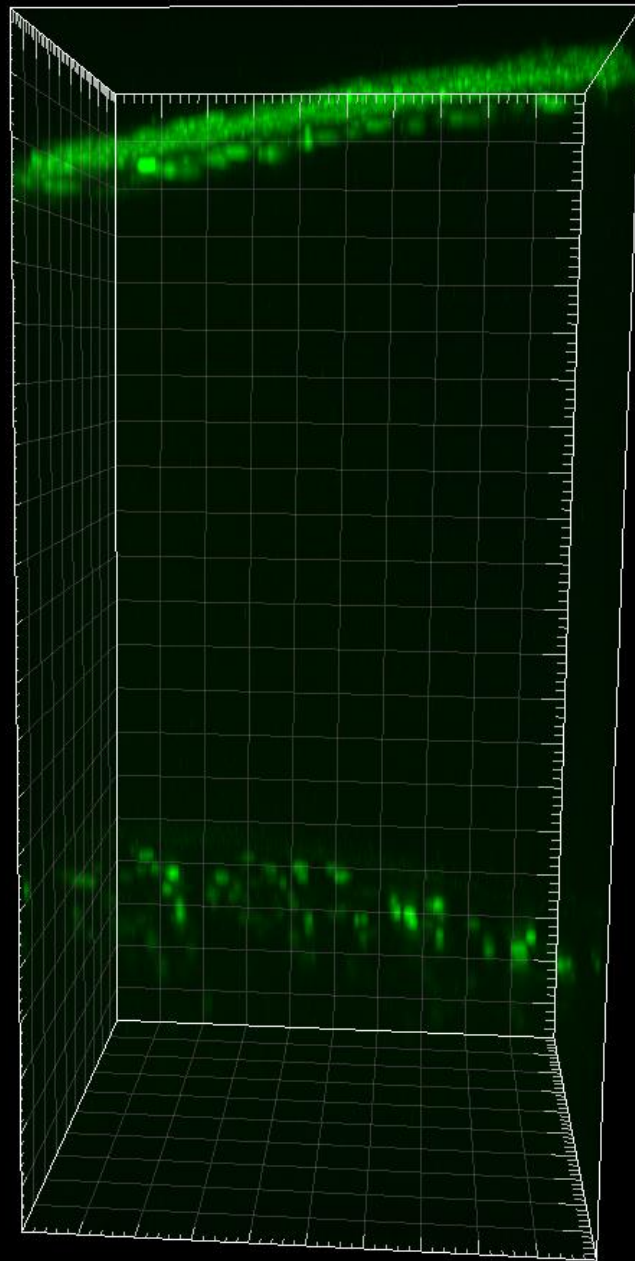
- Pulsed laser needed, thus
- Expensive
- Light path needs constant maintenance
(no glass fiber possible)
- Separation of neighboring fluorochromes
more difficult

Application examples



With
Angela Kurz,
AG Sperandio

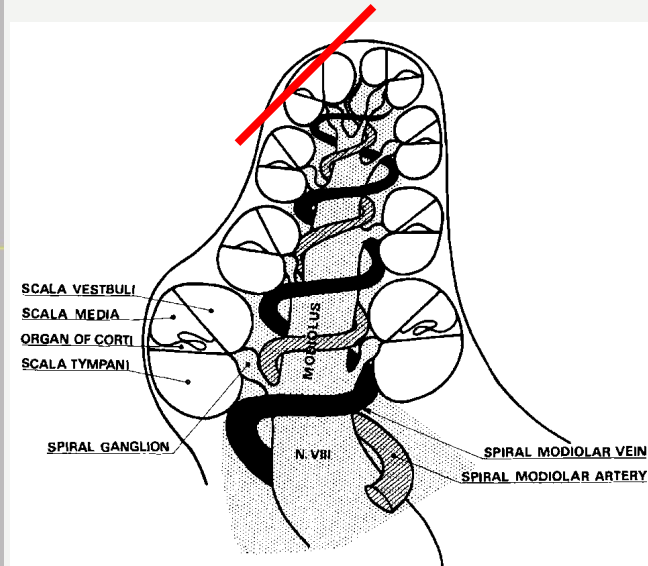
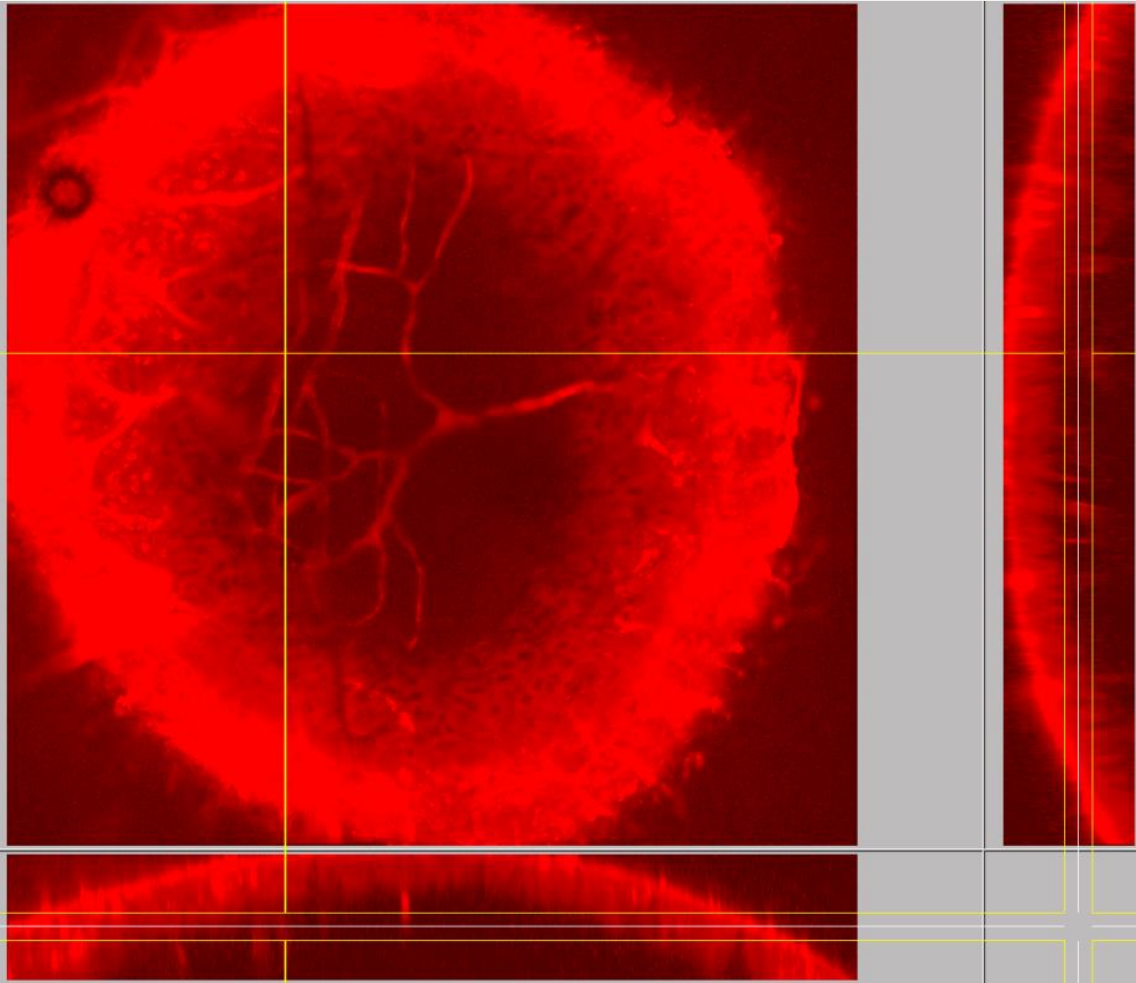
Ly-EGFP mouse
embryo, imaged
through yolk sac

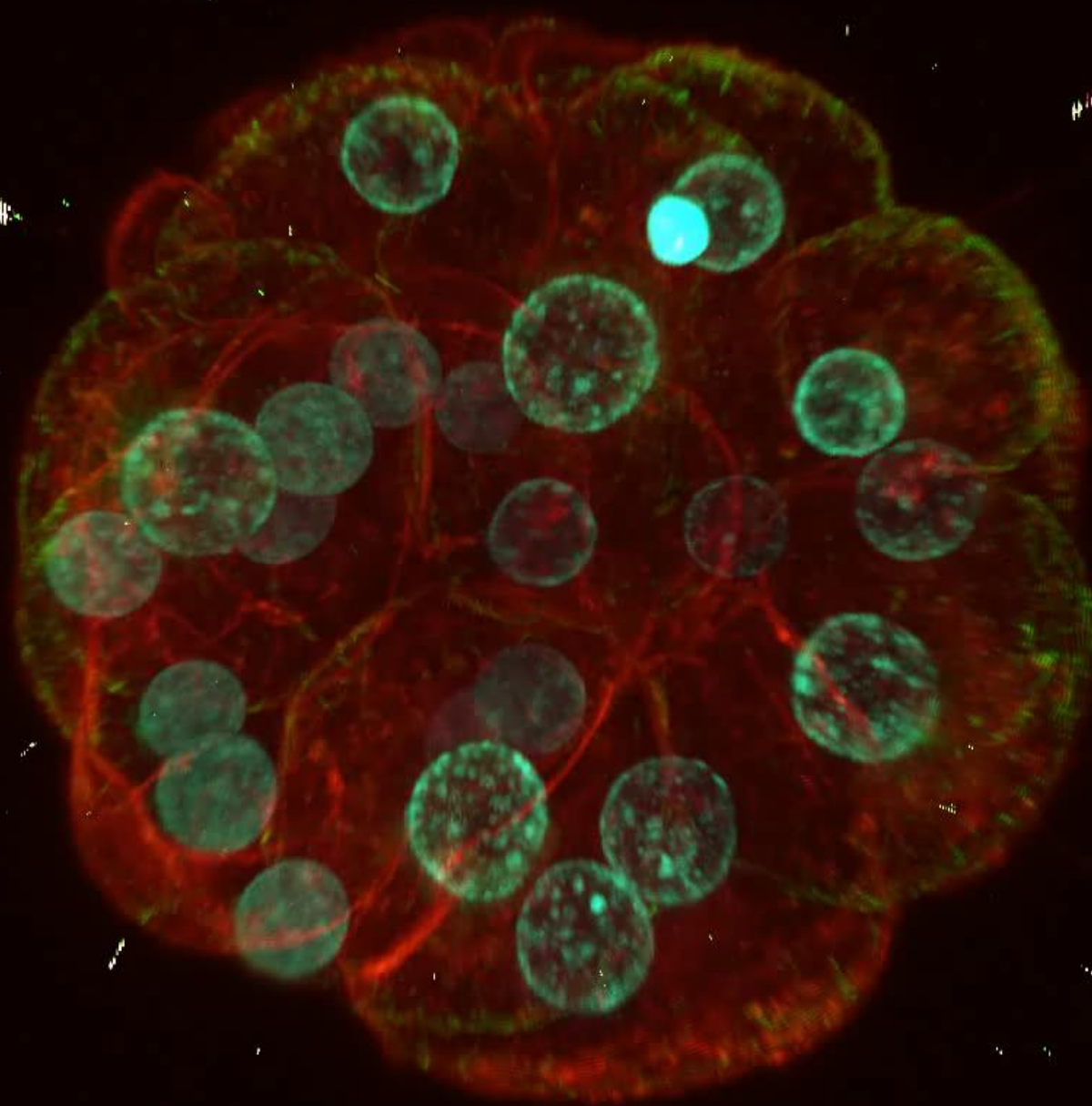


With
Andreas Margraf
AG Sperandio

150 μm

Non-invasive visualization of cochlear microcirculation, with Fritz Ihler, Martin Canis





With Felix
Habermann

Second and Third Harmonic Generation (SHG + THG)



2-photon excited fluorescence

Excited electronic states

$h\nu_1$
 $h\nu_1$
(e.g. 800 nm)

$h\nu_{2PEF}$

Vibrational energy levels

Ground electronic state

SHG

Virtual state

$h\nu_1$
 $h\nu_1$
(e.g. 1275 nm)

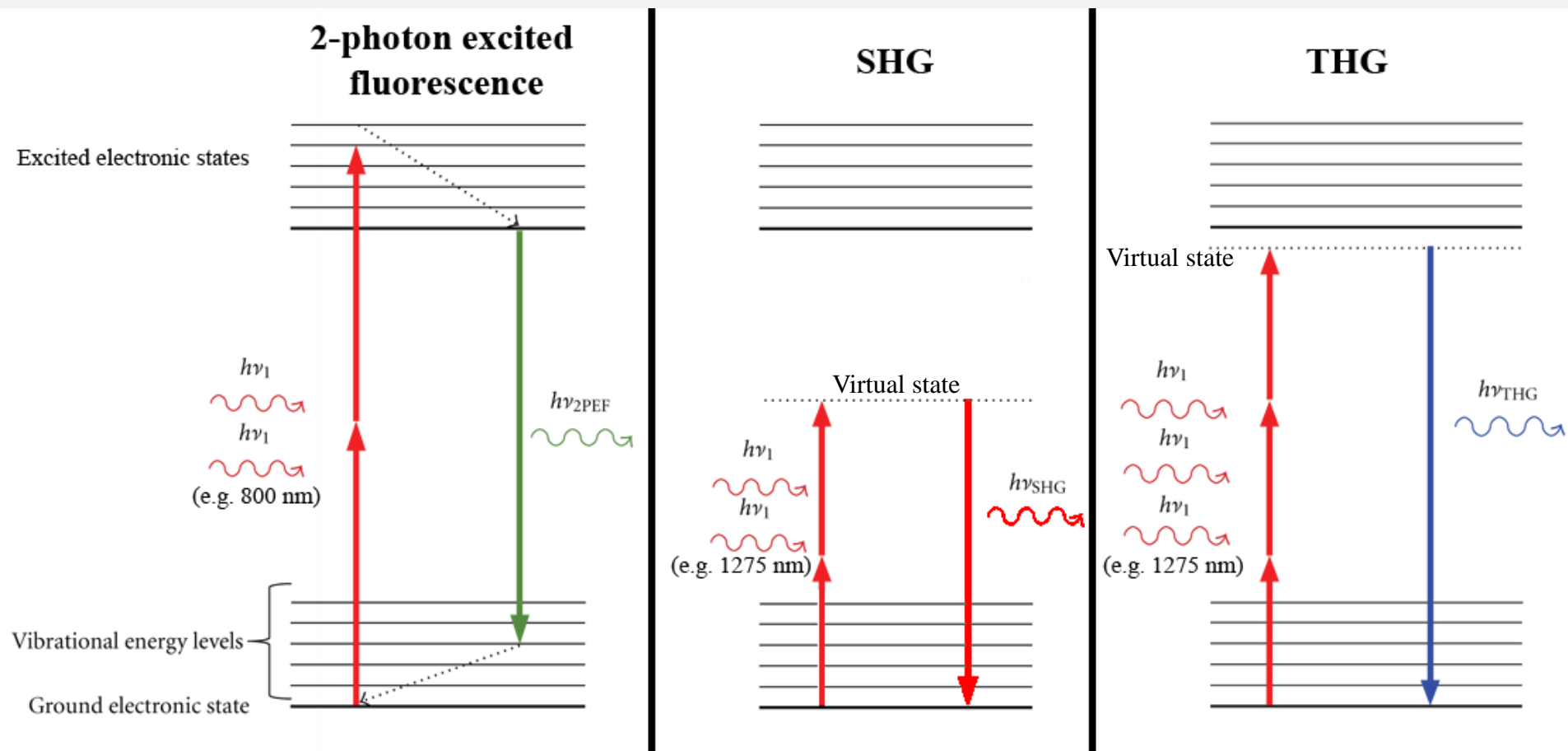
$h\nu_{SHG}$

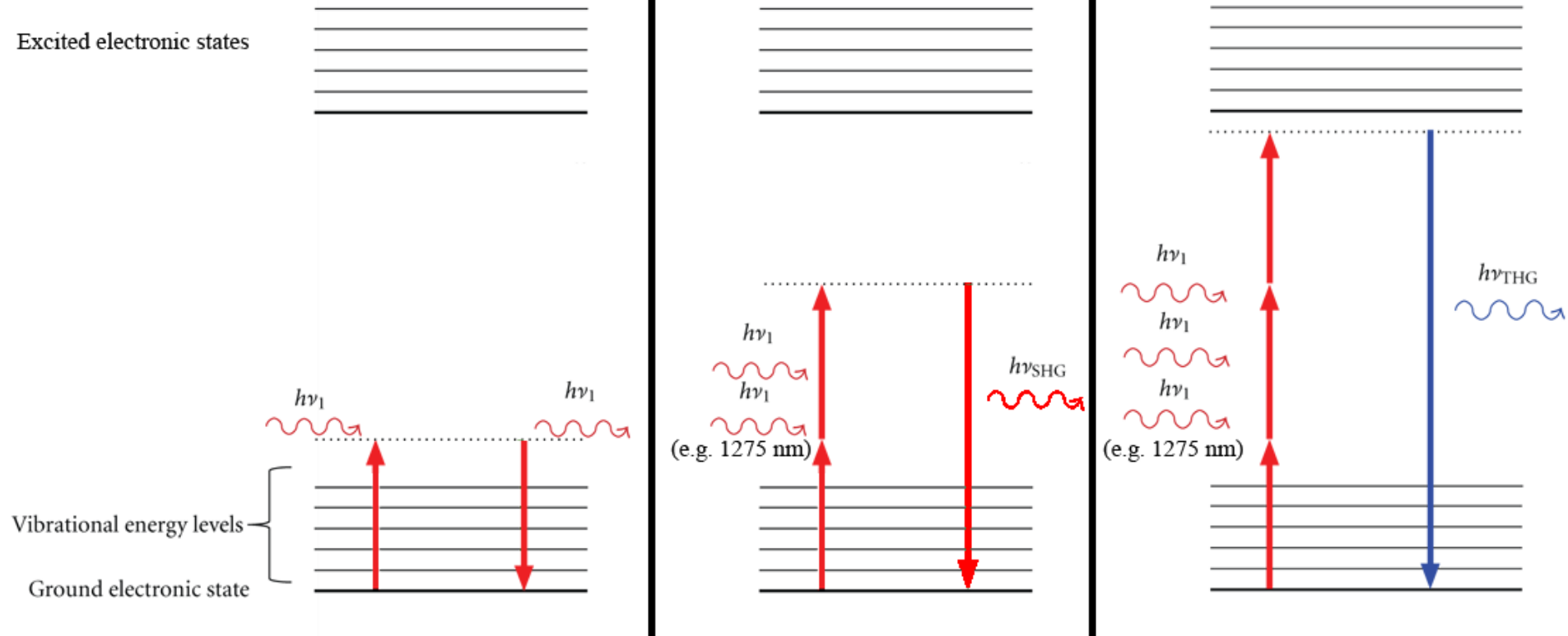
THG

Virtual state

$h\nu_1$
 $h\nu_1$
 $h\nu_1$
(e.g. 1275 nm)

$h\nu_{THG}$



**normal
scattering****SHG****THG**

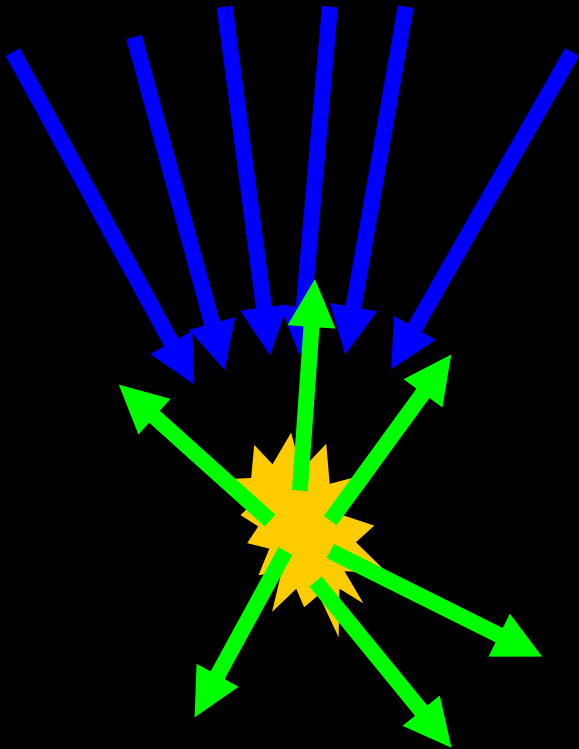
SHG

Two photons in, one out: energy is constant, thus wavelength is exactly halved: 860 nm → 430 nm or 1275 nm → 638 nm

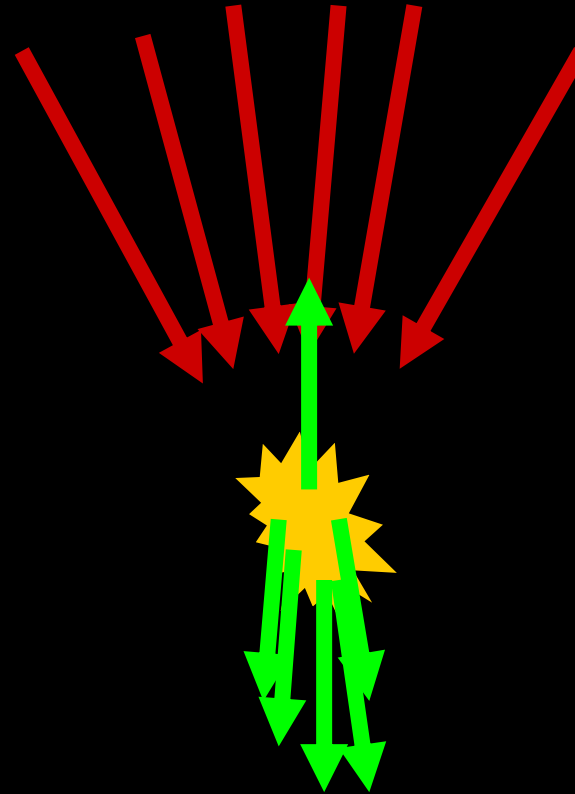
In mammalian soft tissues, generated in collagen fibers and striated muscle myosin (non-centrosymmetric, dense substances)

Label-free, 3D

Fluorescence



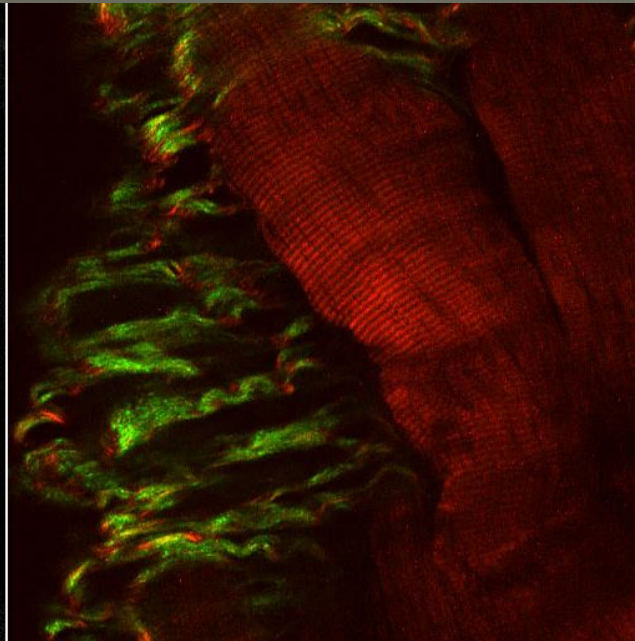
Higher harmonic generation



Forward and backward SHG are not the same



Backward



Forward
(Detected behind the condenser)

- Collagen and myosin both generate stronger SHG-Signals in forward direction than in backward direction. However, the relative backward component is stronger for collagen than for myosin
- Signals are also polarization dependent.

SHG

Two photons in, one out: energy is constant, thus wavelength is exactly halved: 860 nm → 430 nm or 1275 nm → 638 nm

In mammalian soft tissues, generated in collagen fibers and striated muscle myosin (non-centrosymmetric, dense substances)

Label-free, 3D

THG

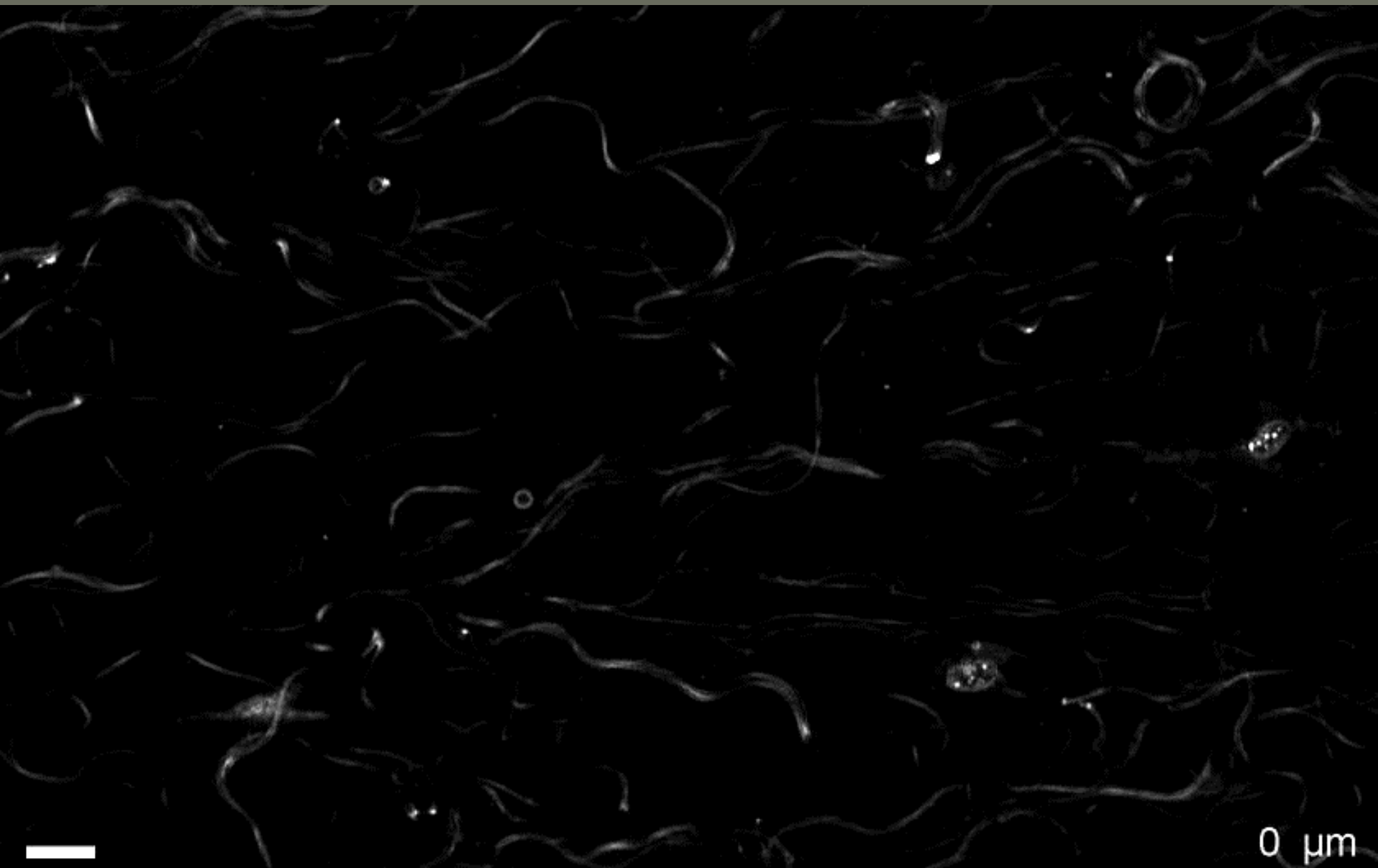
Three photons in, one out: energy is constant, thus wavelength is exactly 1/3: 1275 nm → 425 nm.

Generated at interfaces, e.g. at membranes or refraction index mismatches

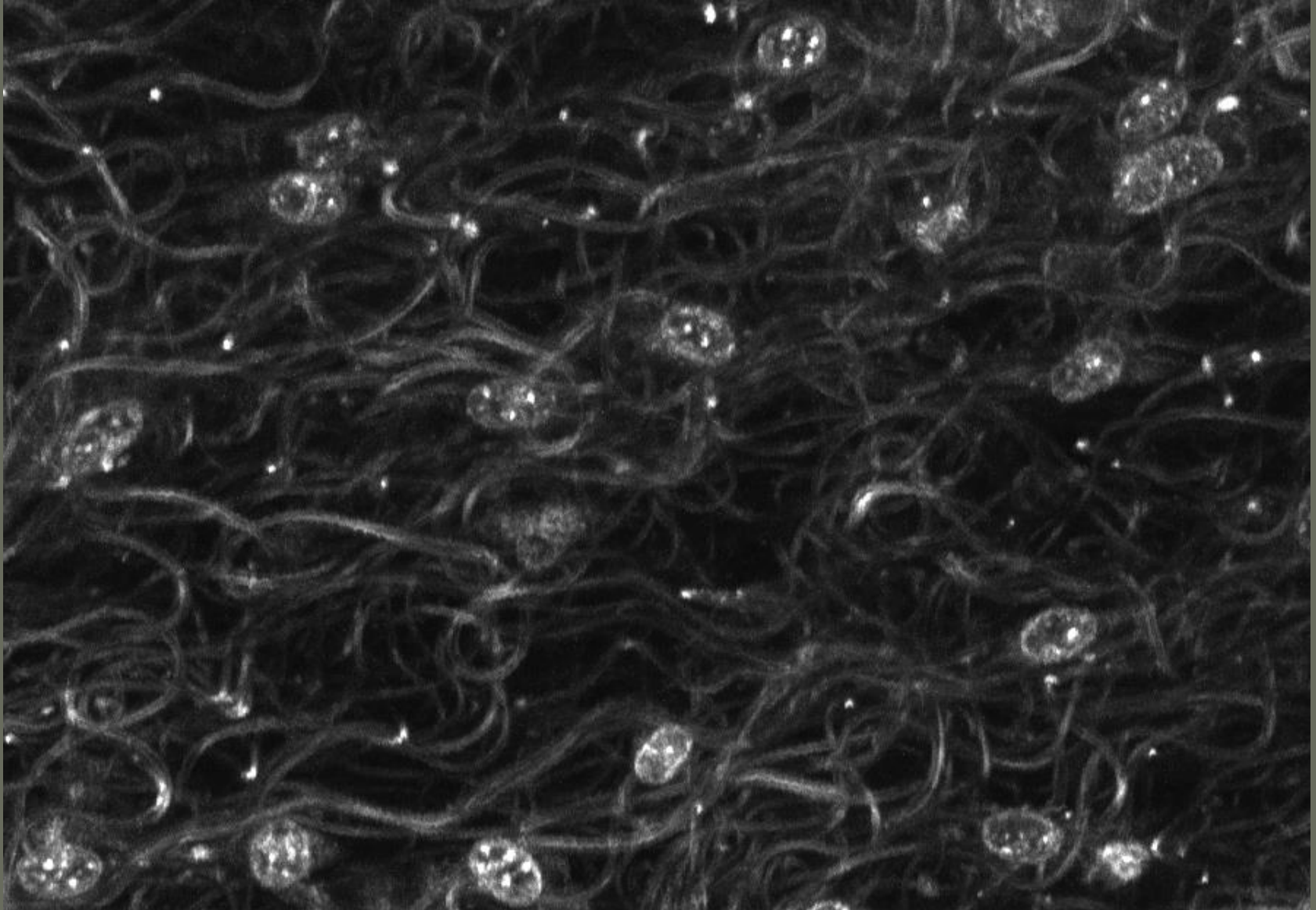
Label-free, 3D

Z-sections

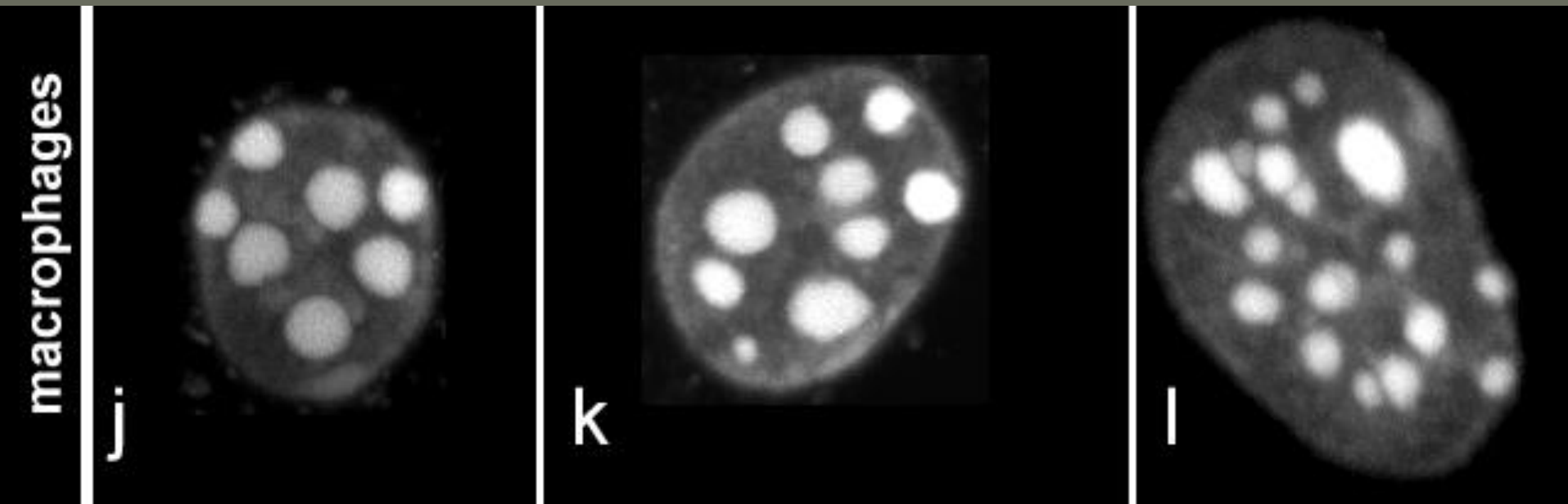
With
Markus Rehberg



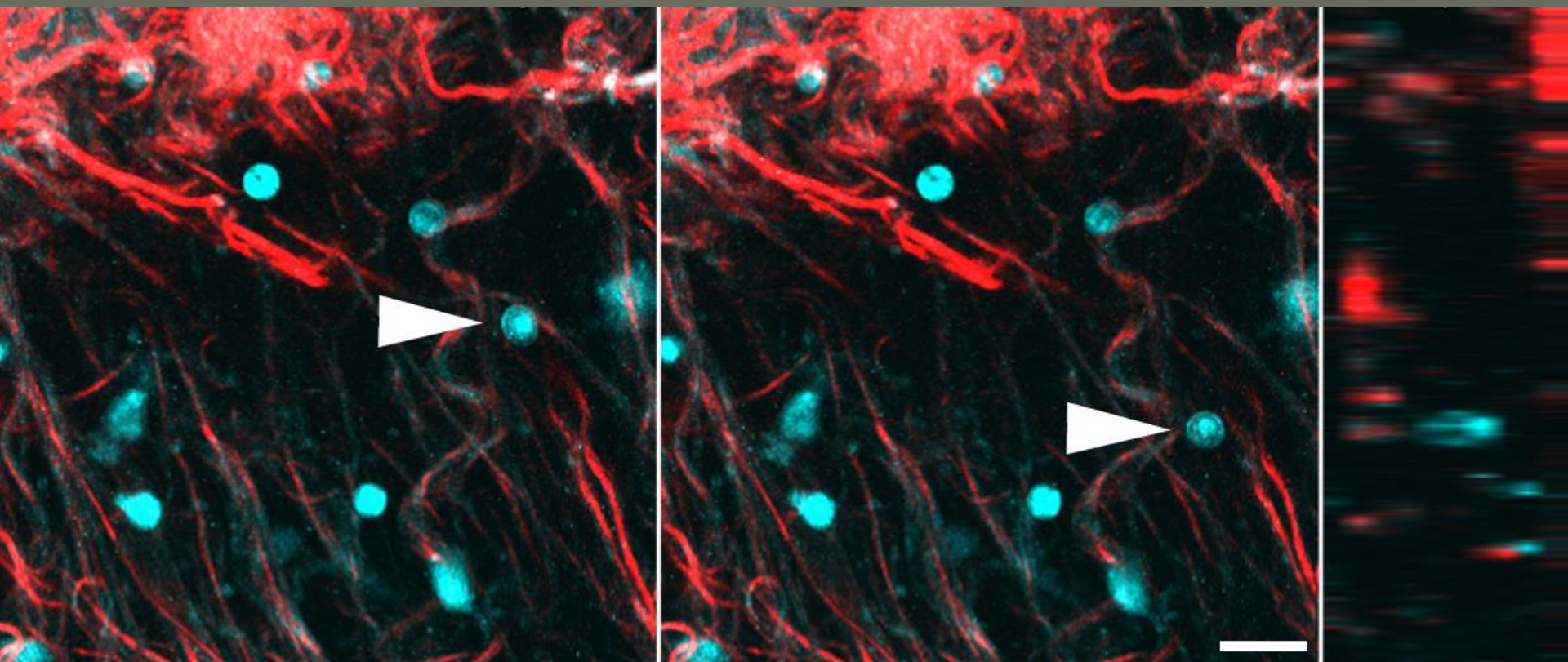
Chromatin structure



Chromocenters in mouse cell nuclei

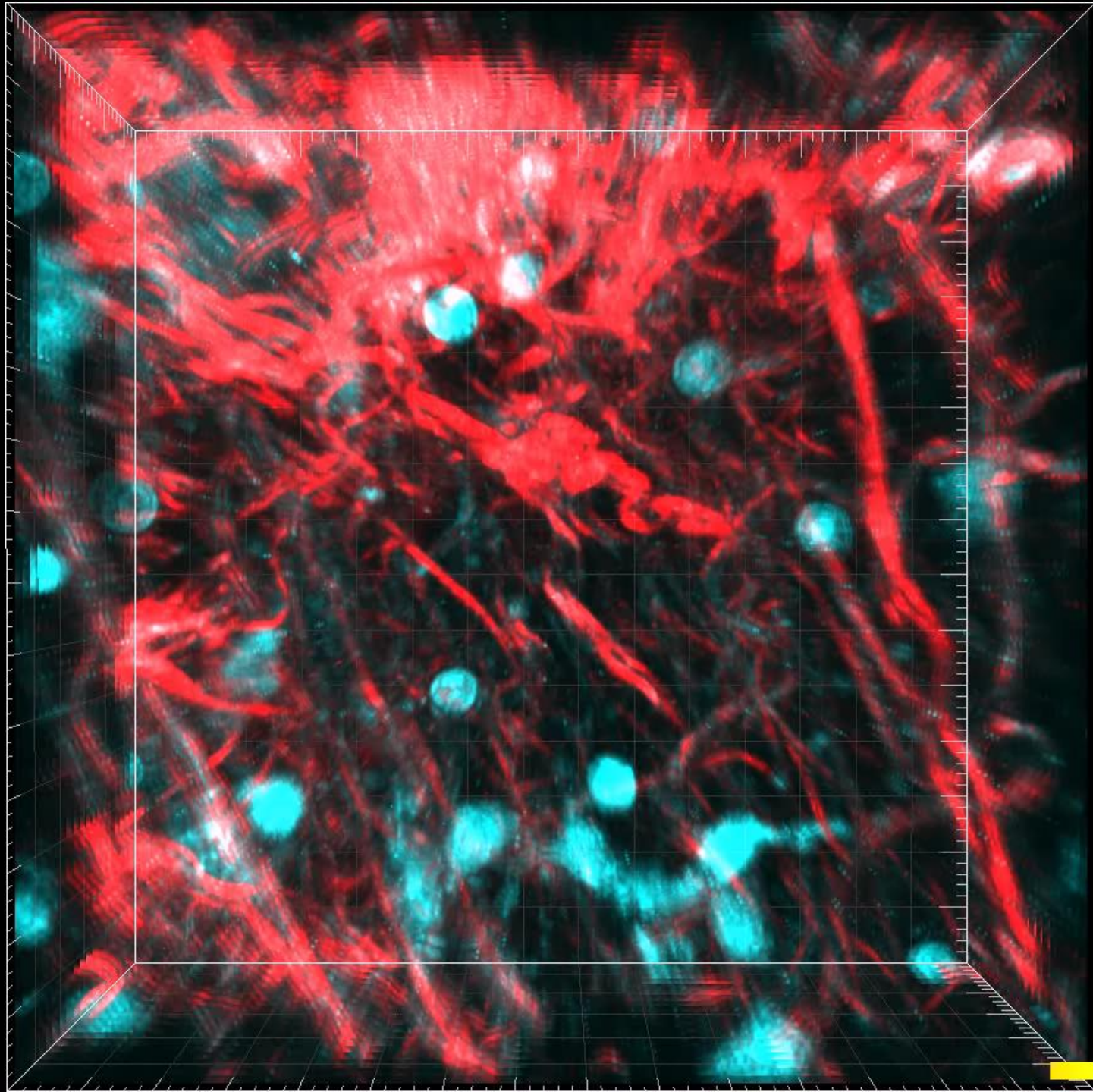


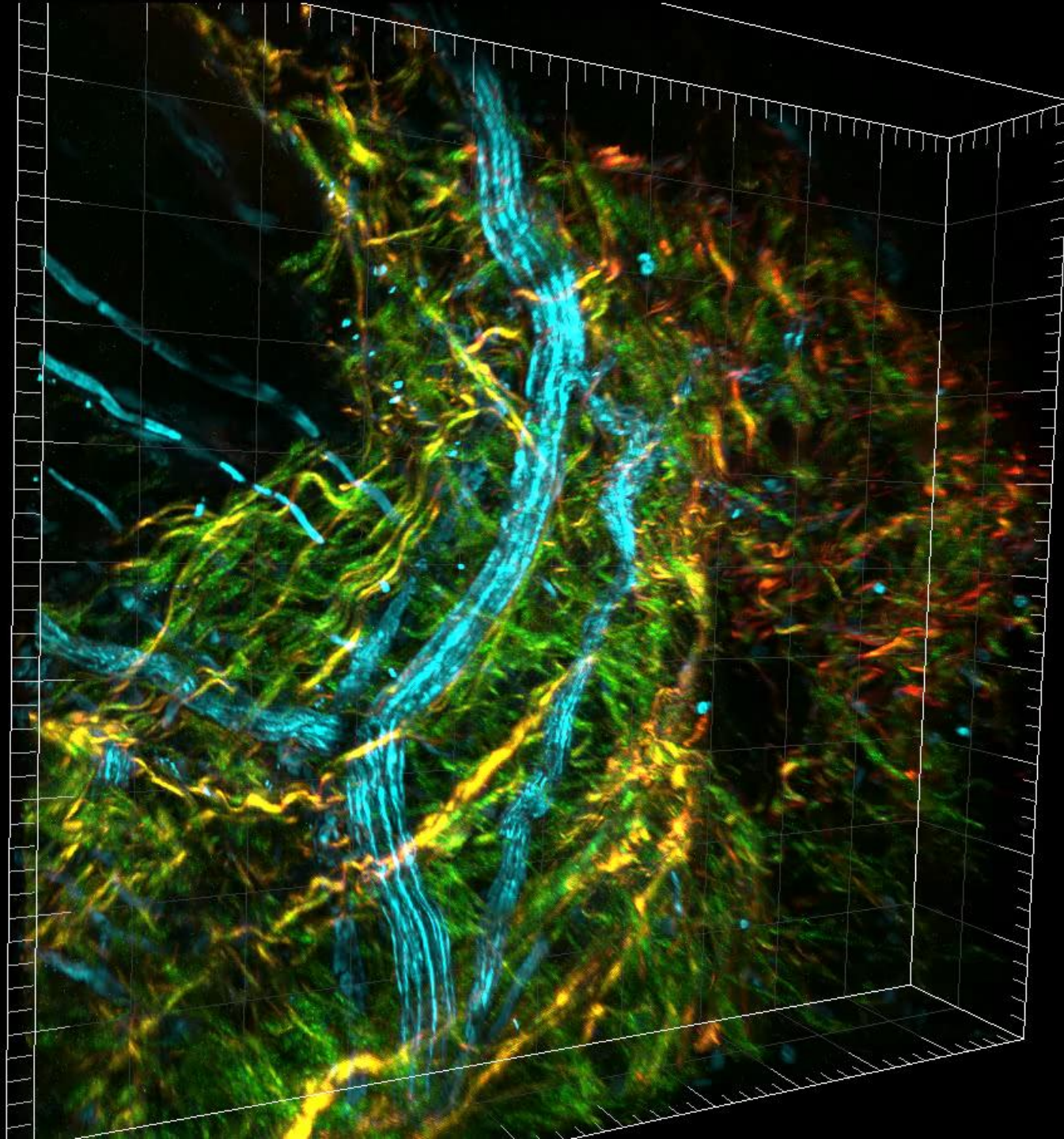
Mayer, R., A. Brero, J. von Hase, T. Schroeder, T. Cremer, and S. Dietzel. 2005. Common themes and cell type specific variations of higher order chromatin arrangements in the mouse. *BMC Cell Biol.* 6:44.



With
Markus Rehberg

20 μm
0000:00:00.000

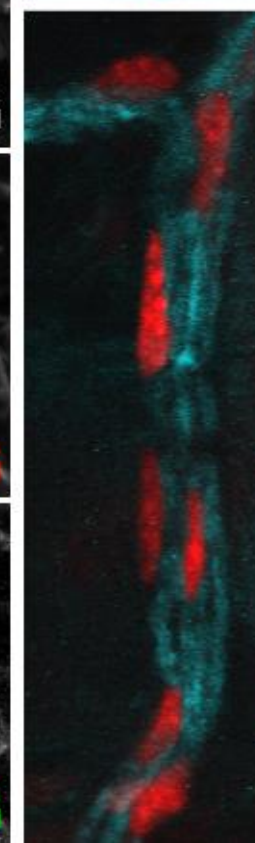
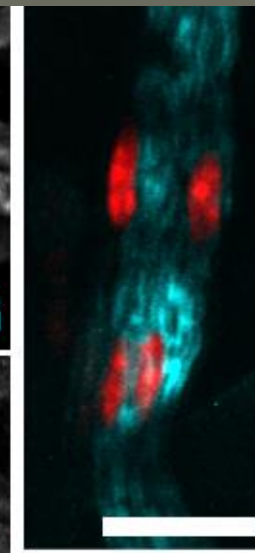
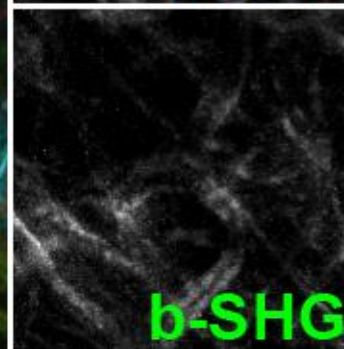
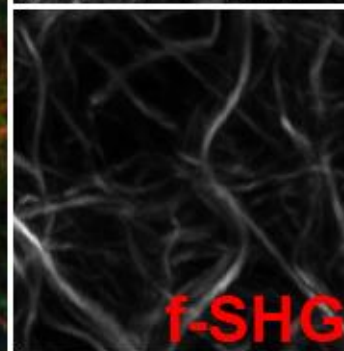
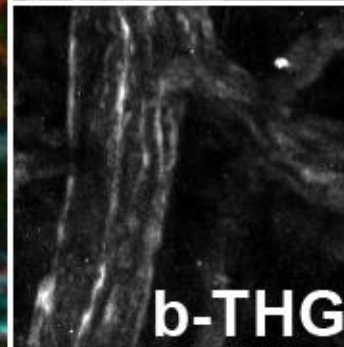
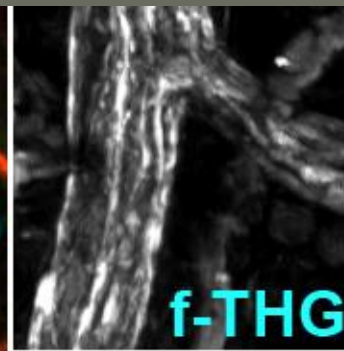
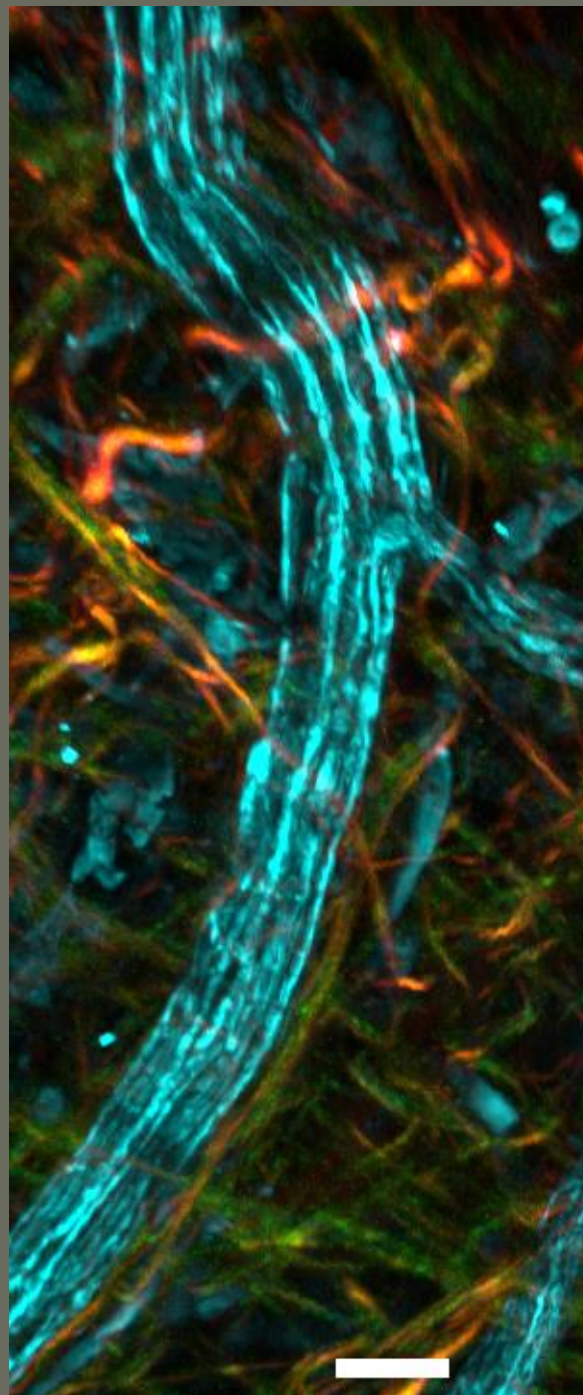




F-THG $^{\Delta\times}$
B-SHG $^{\Delta\times}$
F-SHG $^{\Delta\times}$

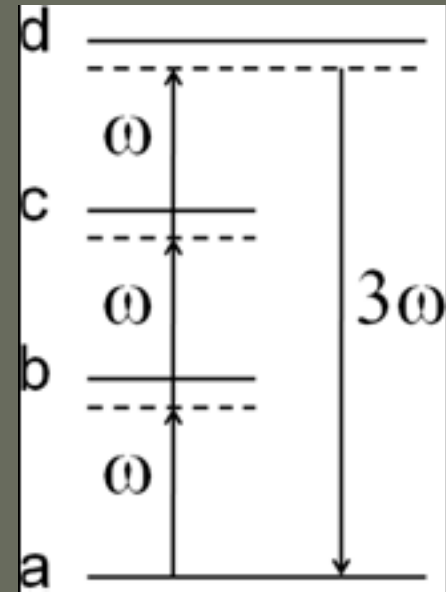
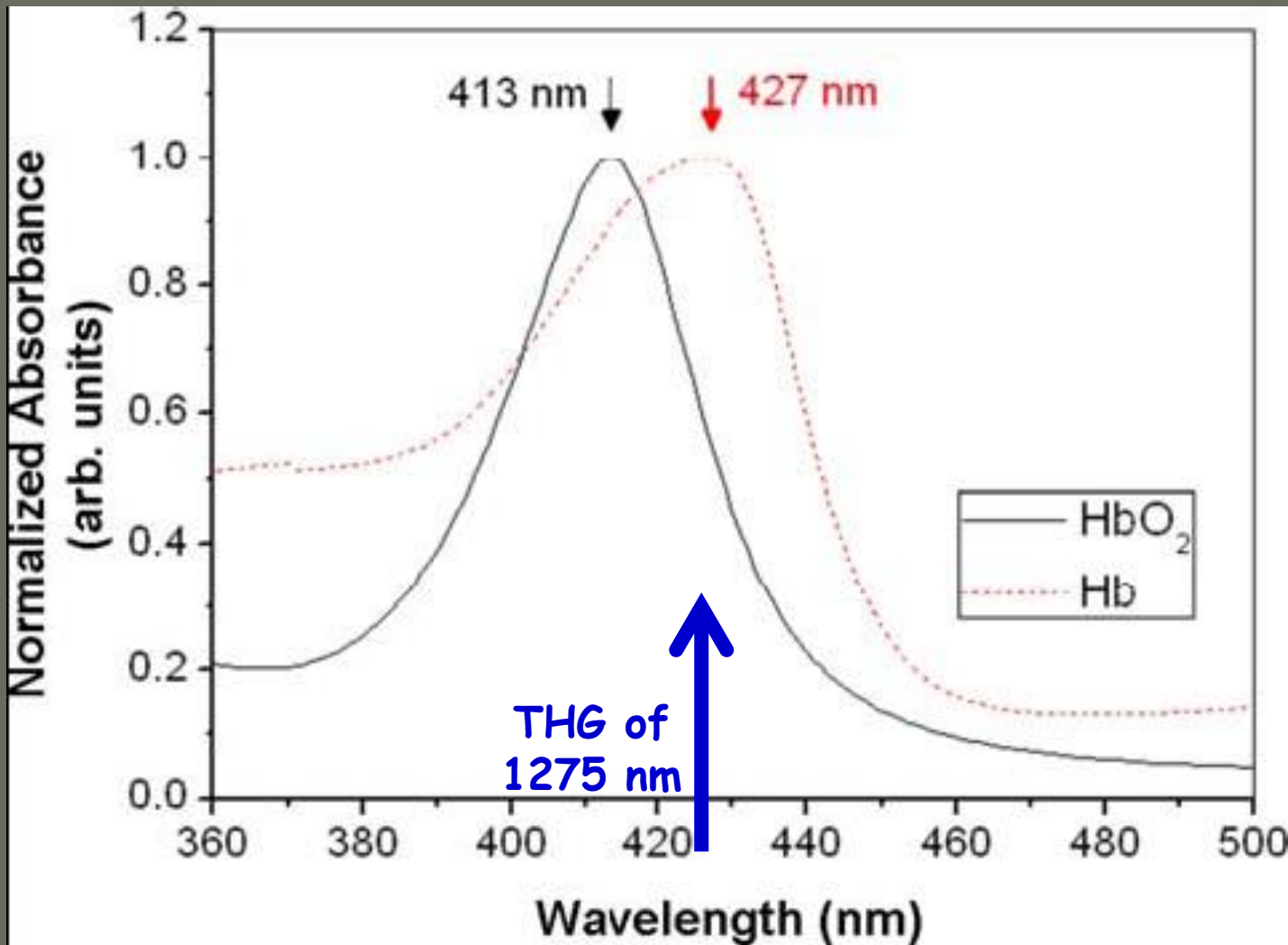
50 μm

f-THG = b-THG
but
f-SHG \neq b-SHG



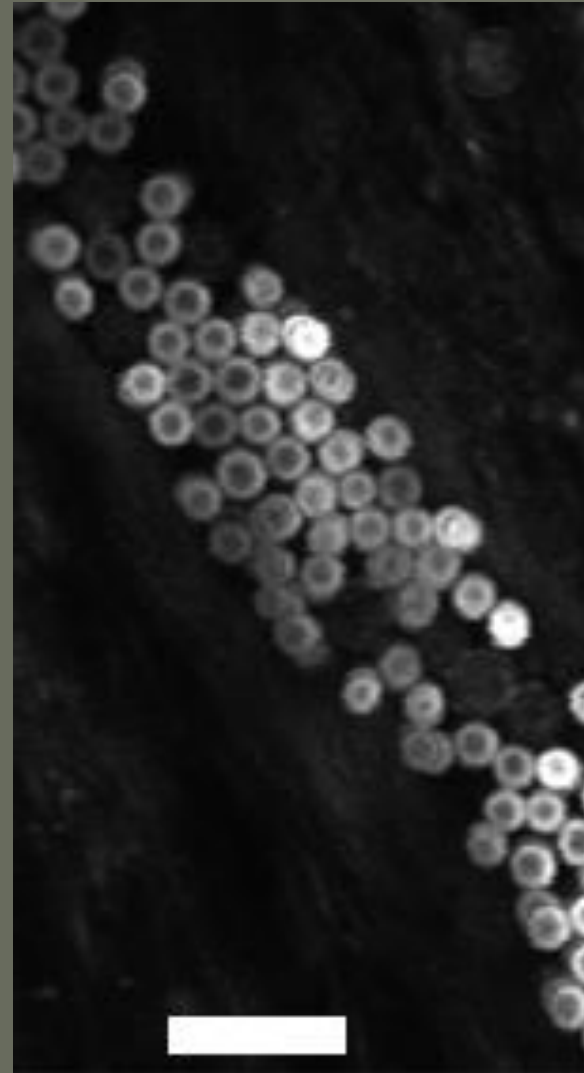
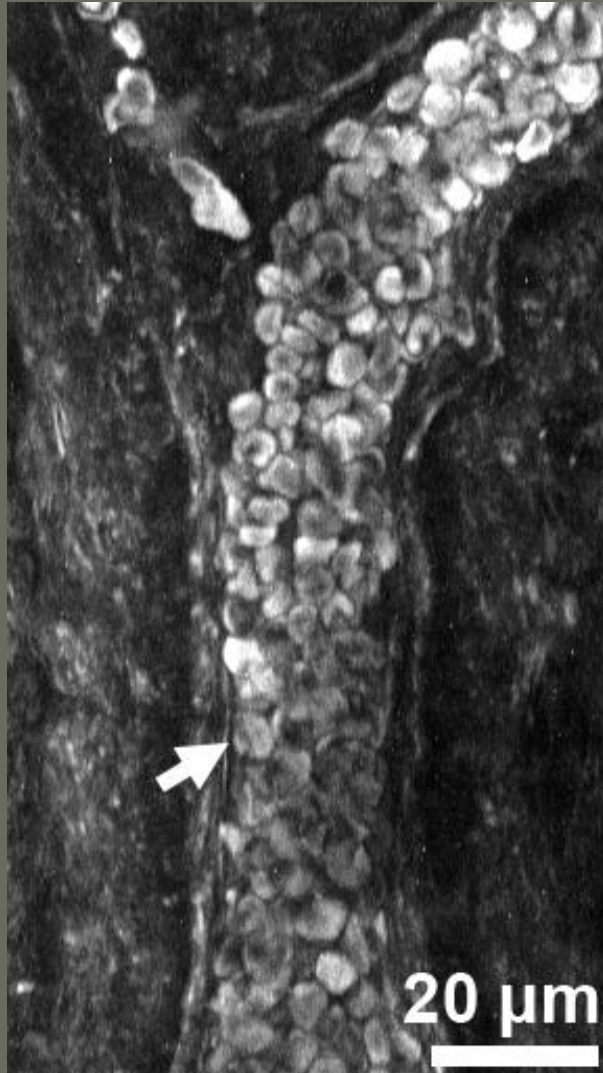
Erythrocytes

Resonance enhancement of THG by hemoglobin

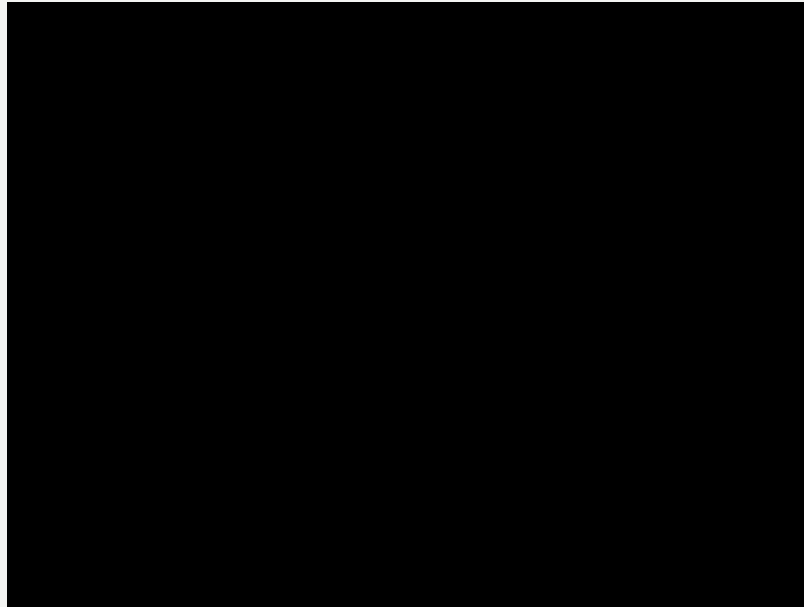


Erythrocytes

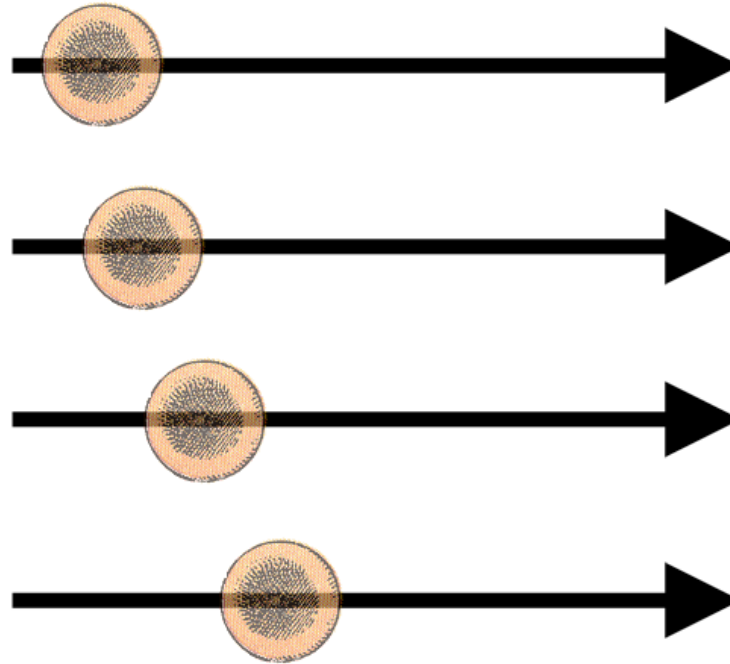
Hemoglobin causes resonance enhancement THG



Fast imaging in capillaries (10 fps)



Line scan principle



Result:

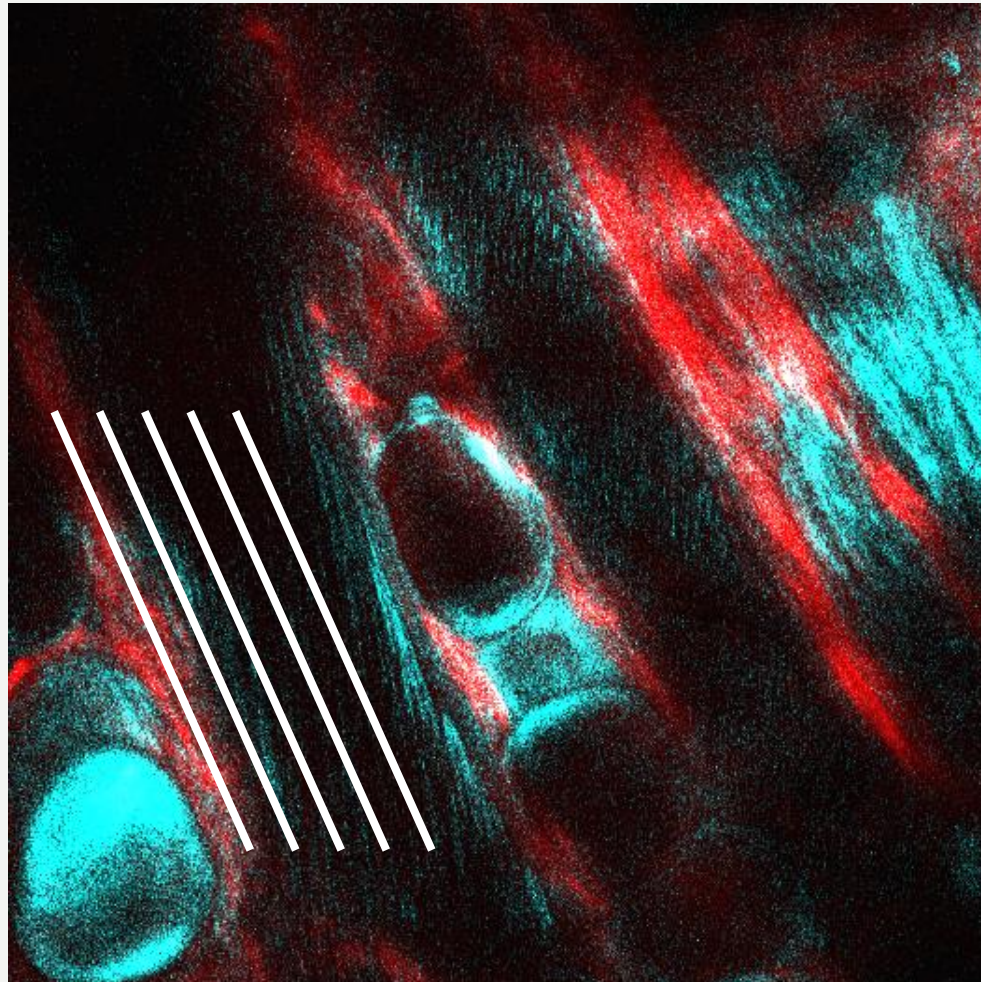


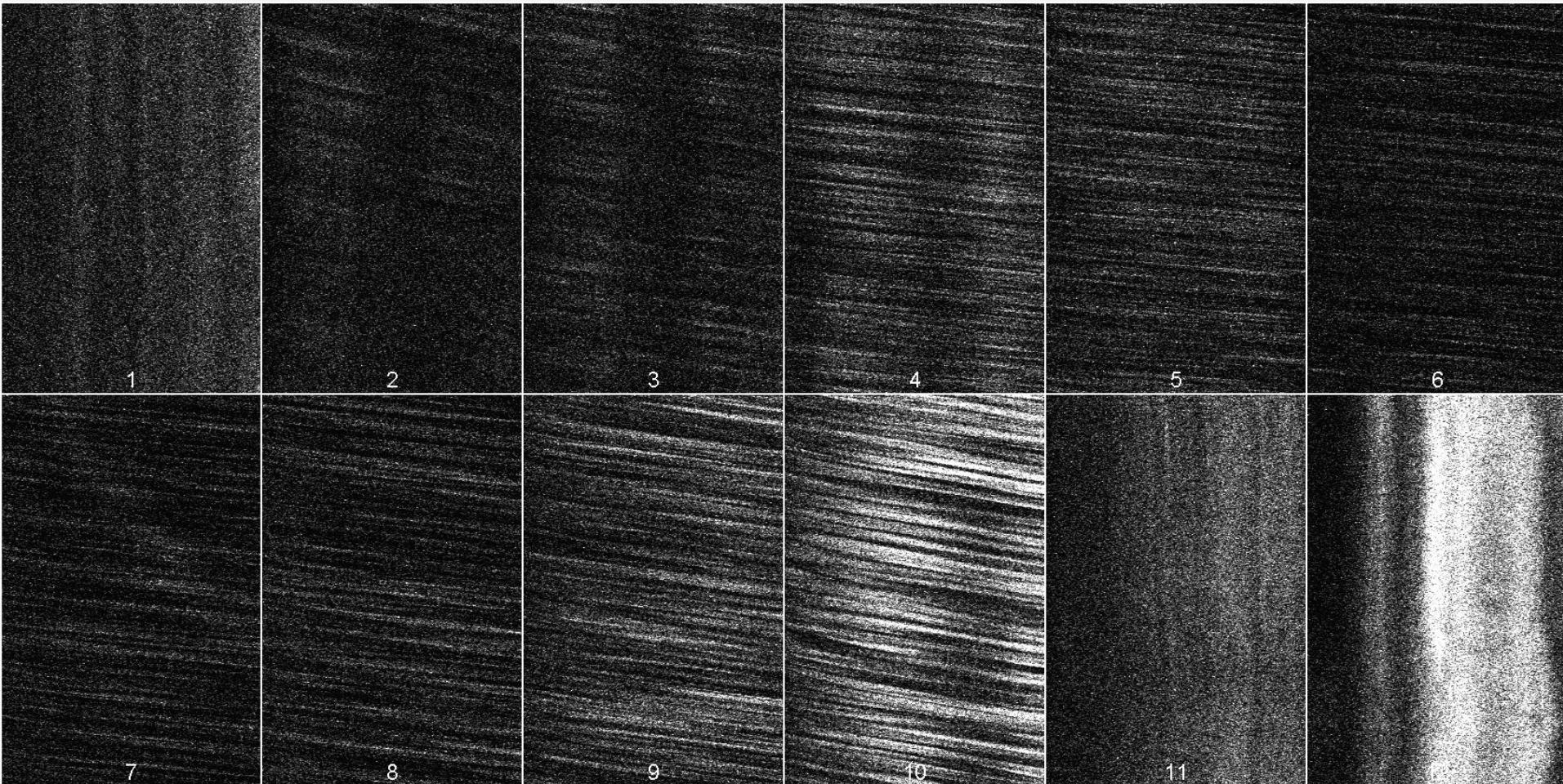
Line scan in the mouse ear



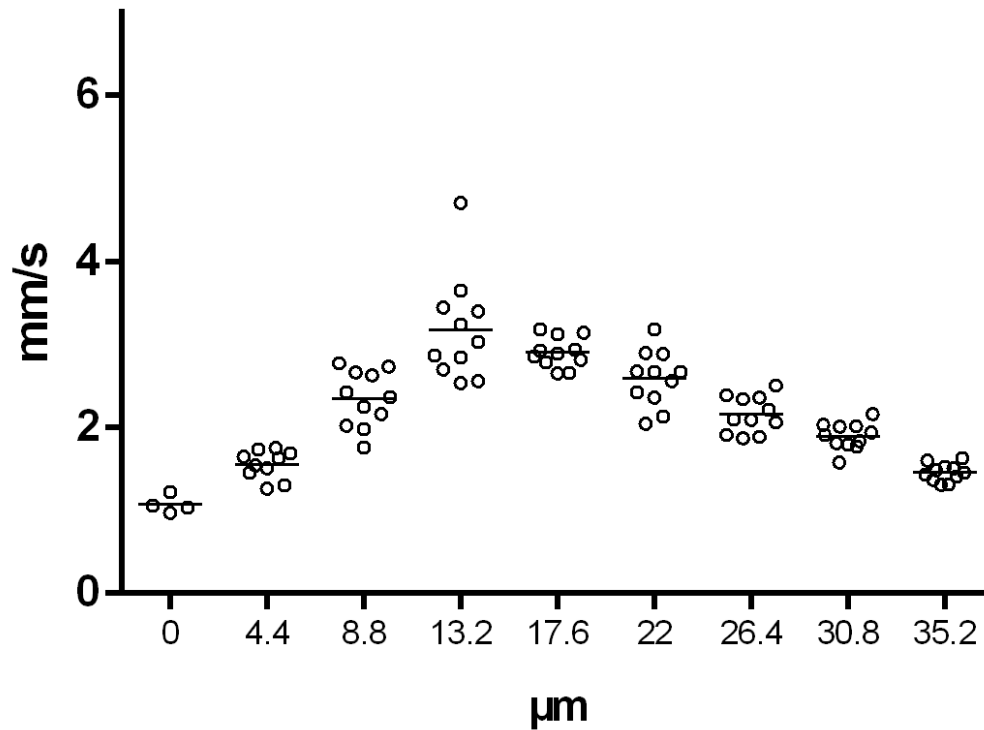
THG blood flow measurements in the ear allows to measure label-free *and* non-invasively.

Scans with shifted line

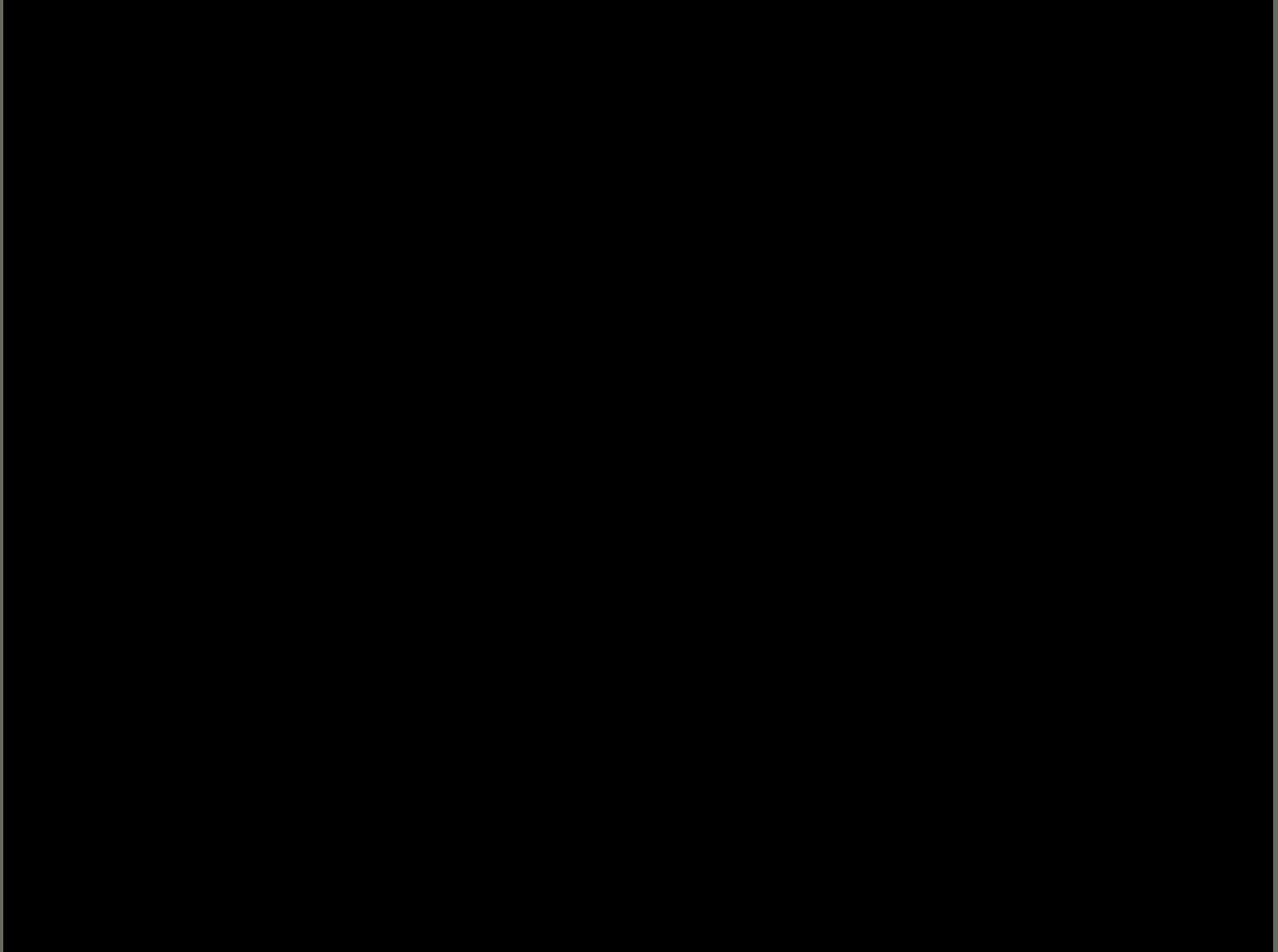


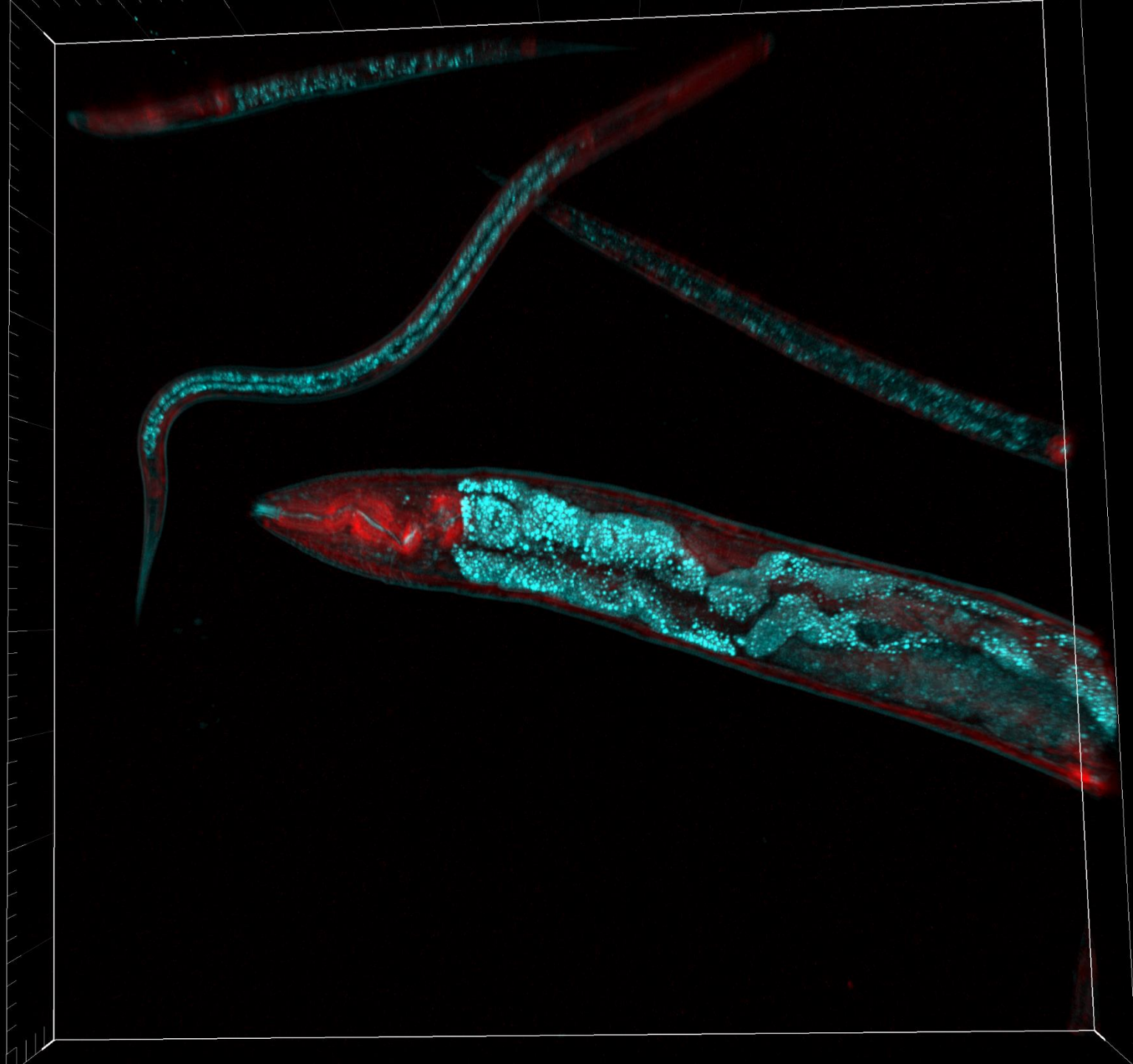


Blood flow in a mouse ear venule



C. elegans

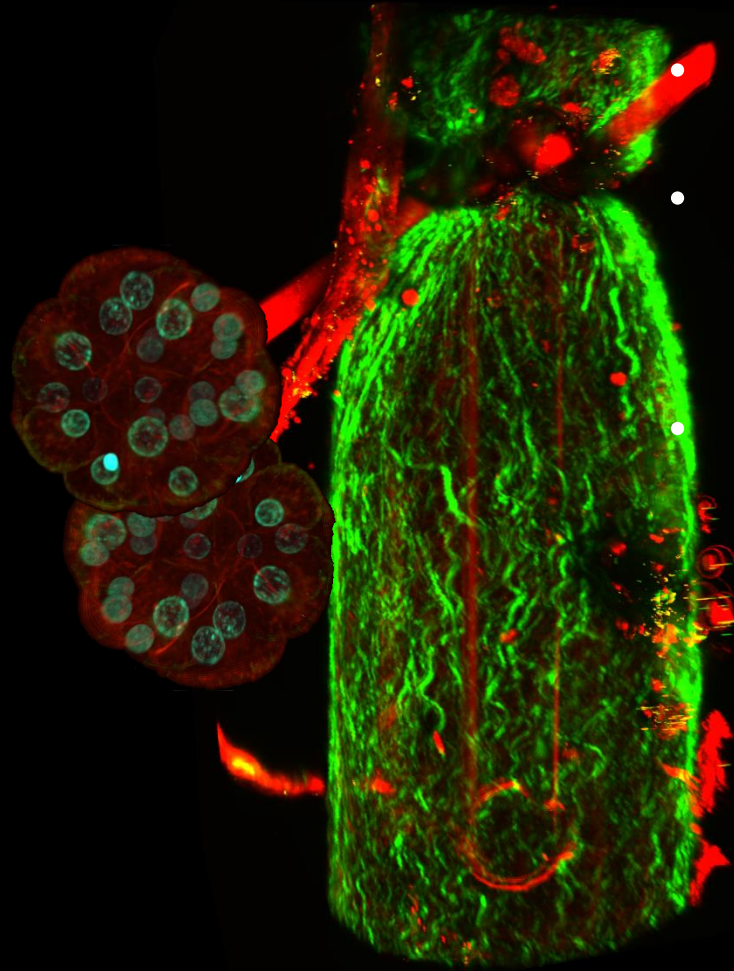




50 μ m

Collaborators

- Fabian Kellner,
Michael Schubert,
Ulrich Pohl
- Markus Rehberg,
Fritz Krombach
- Angela Kurz,
Andreas Margraf
Markus Sperandio



- Fritz Ihler, M. Canis
- Jan Horstkotte,
Tilman Ziegler,
Christian Kupatt, KUM
- Felix Habermann,
LMU VetMed